

# Detection of Biothreat Agents Using a Multiplexed PCR Test for the BIOFIRE® SPOTFIRE® Instrument

Jason Nielson<sup>1</sup>, PhD , Karol Wright , Karol Poloniec , Diana Sanchez , Chris Genin, Katie Lakman, Nicholas Duclos, Whitney Brownlee, Ryan Pack, Stelian Pop, Emily Kress, Ashley Bates, Marianne Kim, PhD  
<sup>1</sup>jason.nielson@biofiredefense.com  
BioFire Defense, LLC, Salt Lake City, Utah, USA

## ABSTRACT

BioFire Defense has provided PCR-based testing capabilities to military personnel and first-responders for the detection of biothreat pathogens, including the Joint Biological Agent Identification and Diagnostic System (JBAIDS), RAZOR® Mk II, and Next Generation Diagnostic System (NGDS) FILMARRAY® 2.0 platforms. These portable systems have provided timely and reliable answers to support critical decision making and countermeasures related to biological warfare agents. The ability to reliably detect biothreat pathogens with the state-of-the-art BIOFIRE® SPOTFIRE® System represents the next generation of biothreat detection capabilities provided by BioFire Defense.

The BioThreat Panel was developed for use on the FILMARRAY 2.0 instrument for detection of bacterial and viral pathogens, and toxin-encoding genes, directly from environmental samples. The BioThreat Panel targets: *Bacillus anthracis*, *Brucella melitensis*, *Burkholderia mallei/pseudomallei*, *Coxiella burnetii*, *Francisella tularensis*, *Rickettsia prowazekii*, *Yersinia pestis*, Eastern equine encephalitis virus, *Marburg Marburgvirus*, *Orthopoxvirus* spp., variola virus, Venezuelan equine encephalitis virus, Western equine encephalitis virus, *Zaire ebolavirus*, and toxin-encoding genes from *Clostridium botulinum* (botulinum toxin) and *Ricinus communis* (Ricin toxin).

The study presented demonstrates that the BioThreat Panel produces similar sensitivity on the next generation, state-of-the-art SPOTFIRE instrument to FILMARRAY 2.0 performance.

Figure 1. The BIOFIRE SPOTFIRE Instrument

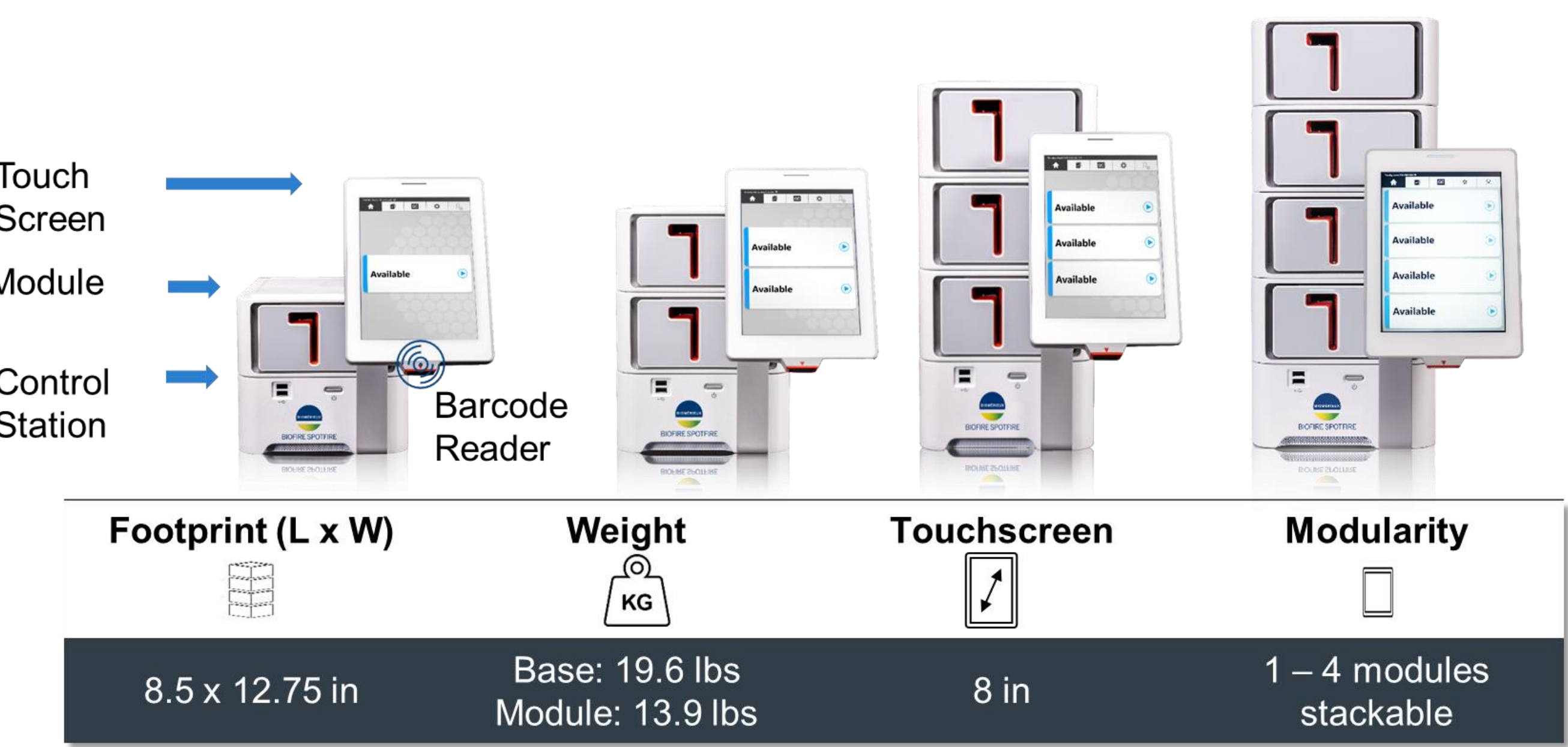


Figure 2. The BIOFIRE Pouch



The SPOTFIRE platform (Figure 1), is a rapid multiplex diagnostic system that can identify many viral, bacterial, fungi, or protozoan pathogens simultaneously. The SPOTFIRE platform is compatible with the existing BIOFIRE Panel consumable format. Following modifications to the instrument instructions governing a pouch run and modifications to the software required to analyze pouch runs, existing BIOFIRE Panels can be compatible with the SPOTFIRE system, achieving run times of ~60 minutes without any changes to reagent chemistry.

The SPOTFIRE System incorporates automated sample extraction (Figure 2A), nucleic acid purification (Figure 2B and 2C), reverse-transcription and nested multiplex first-stage PCR (Figure 2D and 2E) and individual second-stage PCR reactions in a PCR array (Figure 2G) to evaluate multiple targets in a single run (Figure 2). With minimal hands-on time, a comprehensive result is returned.

## STUDY DESIGN

**OVERALL DESIGN:** The study was conducted in two phases, limit of detection (LoD) estimation and LoD confirmation. Testing was performed in phosphate-buffered saline on the SPOTFIRE and FILMARRAY 2.0 instruments in parallel.

**ANALYTE SELECTION:** Live/inactive organisms or genomic DNA were desired. Four analytes: *Eastern equine encephalitis virus*, Variola virus, *Clostridium botulinum*, and *Coxiella burnetii*, did not have material available at a Biosafety Level 2 or below. Synthetic templates were used in place of organism or genomic DNA for these target analytes.

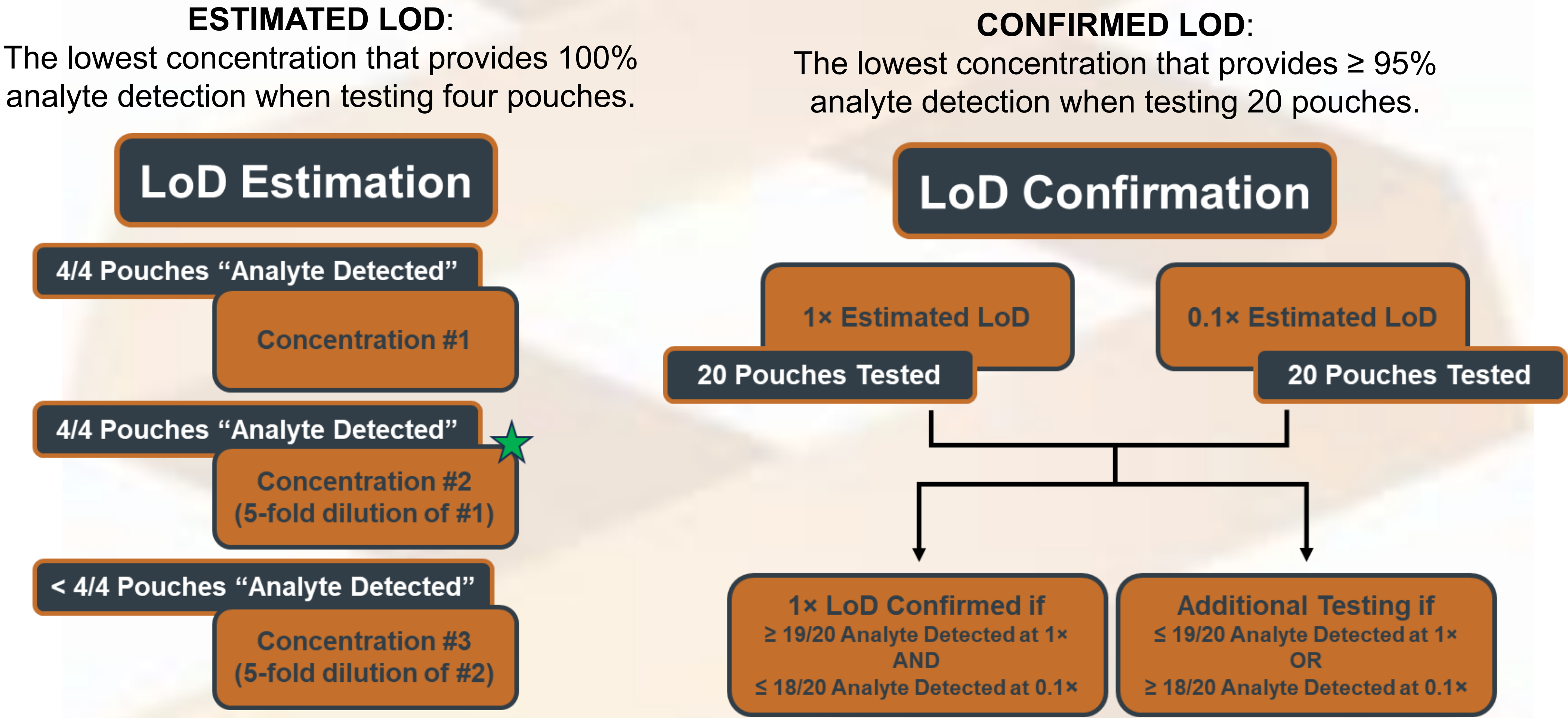


Table 1. Analytes Tested for Each Target on the BioThreat Panel<sup>1</sup>

|          | Target Analyte                                 | Analyte Strain <sup>1</sup>          | Vendor ID <sup>2</sup> | Type of Analyte                | Targeted Assays                | Analyte Interpretation                      |
|----------|--|--------------------------------------|------------------------|--------------------------------|--------------------------------|---|
| Bacteria | <i>Bacillus anthracis</i>                      | Ames                                 | AGD 1331               | Inactivated                    | Chromosome Element, pXO1, pXO2 | <i>Bacillus anthracis</i>                   |
|          |  | Sterne 34F2                          | NR-1400                | Live                           | Chromosome Element, pXO1       | Bacillus spp.                               |
|          | <i>Brucella melitensis</i>                     | 16M                                  | AGD 0074               | Inactivated (Formalin)         | BRT2, BRT4                     | <i>Brucella melitensis</i><br>Brucella spp. |
|          | <i>Burkholderia pseudomallei</i>               | MSHR 146                             | AGD 1595               | Inactivated (Formalin)         | Burk2, Burk8                   | <i>Burkholderia mallei/pseudomallei</i>     |
|          | <i>Coxiella burnetii</i>                       | NA                                   | CHEM-GBK-0485          | Synthetic Nucleic Acid         | CBT1                           | <i>Coxiella burnetii</i>                    |
|          | <i>Francisella tularensis</i>                  | SCHU S4                              | NR-15753               | Inactivated (Formalin)         | FTT2, FTT3                     | <i>Francisella tularensis</i>               |
| Viruses  | <i>Rickettsia prowazekii</i>                   | Brienl                               | AGD 0079               | Inactivated (Formalin)         | RIK2                           | <i>Rickettsia prowazekii</i>                |
|          | <i>Yersinia pestis</i>                         | CO92                                 | NR-2717                | Genomic DNA                    | YPT1, YPT3                     | <i>Yersinia pestis</i>                      |
|          | <i>Zaire ebolavirus</i>                        | Mayinga                              | NR-31807               | Inactivated (Gamma-irradiated) | EB2                            | Ebola Zaire                                 |
|          | Eastern equine encephalitis virus              | NA                                   | CHEM-RNA-0039          | Synthetic Nucleic Acid - RNA   | EEE01                          | EEE Virus                                   |
|          | <i>Marburg marburgvirus</i>                    | German Voegel                        | NR-31816               | Inactivated (Gamma-irradiated) | Marb2, Marb3                   | Marburg virus                               |
|          |  | Ravn                                 | NR-31819               | Inactivated (Gamma-irradiated) | Marb2, Marb3                   |   |
|          | <i>Orthopoxvirus</i> spp.                      | Modified vaccinia ankara (MVA) virus | NR-1                   | Attenuated Live                | OPX2, Var3                     | Orthopox genus virus                        |
|          | Variola virus                                  | NA                                   | CHEM-GBK-0752          | Synthetic Nucleic Acid         | OPX2                           | Variola virus                               |
|          |  | NA                                   | Var1_BTNIIVD_g1QC      | Synthetic Nucleic Acid         | Var1a, Var1b                   |   |
|          |  | NA                                   | Var3_BTNIIVD_g1QC      | Synthetic Nucleic Acid         | Var3                           |   |
| Toxin    | Venezuelan equine encephalitis virus           | Trinidad                             | AGD 0108               | Inactivated (Formalin)         | VEE-MP2, VEE-RC3               | VEE virus                                   |
|          | Western equine encephalitis virus              | CBA87 Alpha 025                      | AGD 0110               | Inactivated (Gamma-irradiated) | WEE01                          | WEE virus                                   |
|          | <i>Clostridium botulinum</i> (Botulinum toxin) | NA                                   | CHEM-RNA-0031          | Synthetic Nucleic Acid         | BoNTA                          | <i>Clostridium botulinum</i>                |
|          | <i>Ricinus communis</i> (Ricin toxin)          | Castor Bean gDNA                     | NR-44091               | Genomic DNA                    | RCN2                           | <i>Ricinus communis</i>                     |

<sup>1</sup>Analyte strain listed for organism isolates; NA listed for synthetic nucleic acid analytes

<sup>2</sup>Vendor ID or BioFire Defense part number, for synthetic nucleic acid templates

## BACKGROUND

## RESULTS

- Similar (within 5-fold) or improved sensitivity on SPOTFIRE for all analytes, except *Yersinia pestis* (Table 2).
- Decrease in sensitivity towards *Yersinia pestis* on SPOTFIRE expected due to changes in the SPOTFIRE software calling scheme.

Table 2. Confirmed LoD Values on SPOTFIRE and FILMARRAY 2.0

|          | Target Analyte                                 | Analyte Strain <sup>1</sup> | Confirmed LoD (copies/mL) |           |
|----------|--|-----------------------------|---------------------------|-----------|
|          |  |                             | SPOTFIRE                  | FILMARRAY |
| Bacteria | <i>Bacillus anthracis</i>                      | Ames                        | 8.0E+03                   | 8.0E+03   |
|          |  | Sterne 34F2                 | 4.0E+03                   | 4.0E+03   |
|          | <i>Brucella melitensis</i>                     | 16M                         | 1.0E+02                   | 1.0E+02   |
|          | <i>Burkholderia pseudomallei</i>               | MSHR 146                    | 4.8E+02                   | 4.8E+02   |
|          | <i>Coxiella burnetii</i>                       | NA                          | 3.0E+04                   | 3.0E+04   |
|          | <i>Francisella tularensis</i>                  | SCHU S4                     | 2.4E+02                   | 2.4E+02   |
| Viruses  | <i>Rickettsia prowazekii</i>                   | Brienl                      | 8.7E+03                   | 1.7E+03   |
|          | <i>Yersinia pestis</i>                         | CO92                        | 2.2E+03                   | 4.4E+01   |
|          | <i>Zaire ebolavirus</i>                        | Mayinga                     | 3.7E+04                   | 1.8E+05   |
|          | Eastern equine encephalitis virus              | NA                          | 7.4E+03                   | 5.9E+05   |
|          | <i>Marburg marburgvirus</i>                    | German Voegel               | 1.4E+04                   | 1.4E+04   |
|          |  | Ravn                        | 3.2E+04                   | 6.5E+03   |
|          | <i>Orthopoxvirus</i> spp.                      | MVA poxvirus                | 3.0E+03                   | 6.0E+02   |
|          | Variola virus                                  | NA                          | 8.0E+03                   | 4.0E+03   |
|          | Venezuelan equine encephalitis virus           | Trinidad                    | 2.4E+03                   | 1.2E+04   |
|          | Western equine encephalitis virus              | CBA87 Alpha 025             | 4.8E+02                   | 2.4E+03   |
| Toxin    | <i>Clostridium botulinum</i> (Botulinum toxin) | NA                          | 3.1E+05                   | 3.1E+06   |
|          | <i>Ricinus communis</i> (Ricin toxin)          | Castor Bean gDNA            | 4.0E+02                   | 8.0E+02   |

<sup>1</sup>Analyte strain listed for organism isolates; NA listed for synthetic nucleic acid analytes  
Confirmed LoD (copies/mL) are listed for each analyte on each instrument. Cells shaded in blue indicate reduced sensitivity on SPOTFIRE and cells shaded in orange indicate increased sensitivity on SPOTFIRE

## TAKEAWAY

- BIOFIRE systems have been widely adopted by the US Department of Defense and others because they provide:
  - Broad capability (tests for more than 100 common and 30 rare infectious disease pathogens)
  - Ease-of-use
  - Speed
  - Low logistics burden (no cold chain required)
- BIOFIRE SPOTFIRE is the next generation of BIOFIRE system
  - FDA-cleared Panels for detection of respiratory pathogens are available for SPOTFIRE
  - Additional Panels in development to provide same capabilities as earlier BIOFIRE platforms, such as the NGDS Warrior Panel
- The SPOTFIRE BioThreat Panel will be available December 2025**

**NOTE: The BioThreat Panel is not for diagnostic use**



See all of BioFire's scientific posters by scanning the QR code to access the Scientific Poster Page

**DISCLAIMER:** The conclusions contained herein are those of the authors and should not be interpreted as representing the official policies or endorsements, either expressed or implied, of the Department of Defense or the U.S. Government.

Presented at the Military Health System Research Symposium (MHSRS)  
August 4 – 7, 2025